that NO and guanylyl cyclase were likely endothelial mediator and effector, respectively, responsible for the endothelium-dependent actions of rutaecarpine.<sup>28</sup>

For experiments designed to study the possible roles of Ca<sup>2+</sup> in the actions of rutaecarpine, both removal of extracellular Ca<sup>2+</sup> and treatment with the intracellular Ca<sup>+2</sup> antagonist, 8-(N,N-diethylamino) octvl-3.4,5,-trimethoxybenzoate (TMB-8) (0.1 mM), suggested that influx of extracellular Ca<sup>2+</sup> was the major factor contributing to the action of rutaecarpine.<sup>28</sup> The vasorelaxant effect of rutaecarpine appeared to be largely dependent on extracellular Ca<sup>+2</sup>, as rutaecarpine failed to induce any relaxation in Ca<sup>2+</sup>-free, EGTA-containing medium, indicating the possible involvement of transmembrane Ca<sup>2+</sup> influx. Moreover. pertussis toxin (100 ng/mL) suppressed the relaxation potency of histamine but had no effects on the action of rutaecarpine. 28 Sodium fluoride (NaF; 1, 2, or 3 mM), a G protein activator,<sup>29</sup> attenuated the action of acetylcholine (ACh), but only minimally affected rutaecarpine. 28 1-[6-{[17β-3-Methoxyestra-1,2,3(10)-trien-17yl]amino}hexyl]-1H-pyrrole-2,5-dione (U73122) (1-10 μM), a phospholipase C inhibitor, <sup>30</sup> suppressed the actions of ACh but had little effect on rutaecarpine. 28

Therefore, rutaecarpine induced an endothe-lium/nitric oxide-dependent vasodilatation in rat aorta precontracted by phenylephrine. These responses could be inhibited by the removal of extracelluar Ca<sup>2+</sup> in the medium. This vasodilatation induced by rutaecarpine depended primarily on the influx of Ca<sup>2+</sup> and not on the mobilization of intracellular Ca<sup>2+</sup> ([Ca<sup>+2</sup>]i). Because pertussis toxin, sodium fluoride, and U73122 did not affect rutaecarpine-induced endothelium-dependent vasodilatation, it is speculated that G<sub>i</sub> proteins or G protein-phospholipase C coupling pathways are probably not involved in the action of rutaecarpine on vascular endothelial cells.<sup>28</sup>

## Effect on Platelet Aggregation

In human platelet-rich plasma, rutaecarpine (40-200  $\mu$ M) inhibited aggregation stimulated by a variety of agonists (i.e., collagen, ADP, epinephrine, and arachidonic acid) as shown in Fig. 2.<sup>31</sup> At 120  $\mu$ M, rutaecarpine almost completely inhibited platelet aggregation induced by arachidonic acid (Fig. 2). Furthermore, rutaecarpine also dose-dependently inhibited collagen (10  $\mu$ g/mL)- and ADP (20  $\mu$ M)-induced plate-

let aggregation.<sup>31</sup> However, even at 200 µM, it did not completely inhibit platelet aggregation induced by collagen, ADP, or epinephrine (Fig. 2). The IC<sub>50</sub> values for platelet aggregation induced by collagen, epinephrine, ADP, and arachidonic acid were estimated to be about (µM) 166.2, 64.8, 159.6, and 76.5, respectively. 31 The antiplatelet activity of rutaecarpine (120 µM) was not significantly attenuated by pretreatment with the nitric oxide synthase inhibitors, N<sup>G</sup>-mono-methyl-Larginine (L-NMMA) (100 μM) or N<sup>G</sup>-nitro-L-arginine methylester (L-NAME) (200 µM), or with the guanylyl cyclase inhibitor, methylene blue (100 µM). In addition, rutaecarpine (40-200 µM) did not significantly affect cyclic AMP or cyclic GMP levels in washed human platelets, whereas it (40-200 µM) significantly inhibited thromboxane B2 (TxB2) formation stimulated by collagen (10  $\mu$ g/mL) and thrombin (0.1 U/mL).<sup>31</sup> In a further characterization of whether or not the inhibition of TxB2 formation was due to the inhibition of throm-

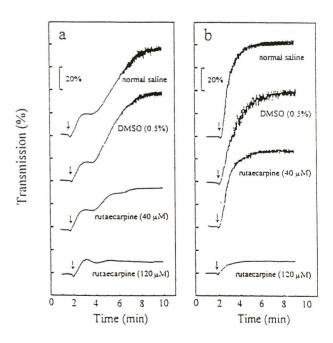


Fig. 2. Typical antiplatelet effect of rutaecarpine on adrenaline (10 μM)- and arachidonic acid (100 μM)-induced aggregation of human platelet-rich plasma. Human platelet-rich plasma was preincubated with normal saline (control), DMSO (0.5%), and rutaecarpine (40 and 120 μM) at 37 °C for 1 min. (a) Adrenaline (10 μM; ↓) or (b) arachidonic acid (100 μM; ↓) was then added to induce platelet aggregation. For the detailed experimental procedure, see Ref. 31.