

行政院國家科學委員會專題研究計畫 成果報告

蘭嶼山欖 (Planchonella duclitan) 葉部所含抗癌活性物
質的分離、純化及結構鑑定

計畫類別：個別型計畫

計畫編號：NSC93-2113-M-038-003-

執行期間：93年08月01日至94年09月30日

執行單位：臺北醫學大學生藥學研究所

計畫主持人：李宗徽

報告類型：精簡報告

處理方式：本計畫可公開查詢

中華民國 94 年 9 月 19 日

中文摘要：

蘭嶼山欖 (*Planchonella duclitan*) 葉部的甲醇萃出液經一系列的分離、純化及結構鑑定，共純化出七個 ursane 類型的三萜酸 (triterpene acids)，分別為 2 α ,3 α ,19 α ,23-tetrahydroxy-13,27-cyclours-11-en-28-oic acid (1), myrianthic acid (2), 2-hydroxyursolic acid (3), ursolic acid (4), pomolic acid (5), rotundic acid (6), and jacoumaric acid (7)，其中化合物 1 為結構上含環丙基 (cyclopropyl) 的新化合物。此外，癌細胞毒殺試驗的結果顯示化合物 4 和 7 對於大腸癌細胞株 HT29 及乳癌細胞株 MCF-7 有顯著的細胞毒性，其半致死濃度範圍為 5.8 \pm 1.4 ~ 6.5 \pm 1.9 μ M。

關鍵詞：蘭嶼山欖、山欖科、葉部、三萜酸、HT29、MCF-7

ABSTRACT

From the methanolic extract of the leaves of *Planchonella duclitan*, 2 α ,3 α ,19 α ,23-tetrahydroxy-13,27-cyclours-11-en-28-oic acid (1), myrianthic acid (2), 2-hydroxyursolic acid (3), ursolic acid (4), pomolic acid (5), rotundic acid (6), and jacoumaric acid (7) were isolated, and their structures were elucidated on the basis of their spectroscopic analysis. Among them, compound 1 was a new cyclopropyl ursane-type triterpene acid. Additionally, compounds 4 and 7 showed significant cytotoxicity toward human colorectal carcinoma cell line HT29 and human breast carcinoma cell line MCF-7 with IC₅₀ values ranging from 5.8 \pm 1.4 to 6.5 \pm 1.9 μ M.

Key Words: *Planchonella duclitan*; Sapotaceae; Leaves; Triterpene acids; 2 α ,3 α ,19 α ,23-tetrahydroxy-13,27-cyclours-11-en-28-oic acid; HT29; MCF-7

INTRODUCTION

Planchonella duclitan (Blanco) Bakhuizen, a tall tree belongs to the family Sapotaceae, is distributed only in the areas of Lanyu Island and South-East Asia.¹ It was used as firewood or for making the decks of ships locally, but not in folk medicines. Recently, it was shown from our preliminary pharmacological experiments that the crude extracts of the leaves of this plant exhibited significant anti-proliferation activities toward breast cancer cell line MCF-7 and liver cancer cell line Hep 3B. The leaves may contain bioactive agents with anti-proliferation activities worthwhile to be investigated phytochemically. Therefore, a series of phytochemical examinations on the leaf extracts of this plant was thus undertaken and has led to the isolation and characterization of seven triterpene acids **1**–**7**. This paper describes the isolation and structural elucidation of the new compound as well as their cytotoxicities.

RESULTS AND DISCUSSION

From the methanolic extract of the leaves of *P. duclitan* seven triterpene acids were identified. The compounds were isolated by a serial separation on Si-gravity column and reversed phase HPLC. Spectroscopic data of 2 α ,3 α ,19 α ,23-tetrahydroxy-12-en-28-oic acid (**2**) were interpreted by comparison with those reported in literatures, and reported as myrianthic acid.² The structure of **3** was determined to be 2 α -hydroxyursolic acid, named as corosolic acid, having been isolated from callus tissue cultures of *Eriobotrya japonica*,³ and *Chaenomeles sinensis*.⁴ Compound **4**, a major component, was obtained as a white powder whose spectral data were consistent with those of ursolic acid, having been isolated from *Ilex paraguariensis*⁵ and *Baeckea gunniana*.⁶ Both compounds **5** and **6** were ursolic acid analogues, and were identified as pomolic acid (**5**), obtained previously from *Sanguisorba officinalis*,⁷ and rotundic acid (**6**), isolated from the root bark of *Mussaenda macrophylla*.⁸ Alkaline hydrolysis of compound **7** afforded corosolic acid (**3**) and *trans-p*-coumaric acid. This result and spectroscopic evidence showed **7** to be jacoumaric acid, which had been isolated from a Chinese Medicine, Goreishi (the feces of *Troglodytes xanthipes*),⁹ and *Leptospermum scoparium*.¹⁰

Compound **1**, a white powder, had a molecular formula of C₃₀H₄₆O₆ based on the results of HRFABMS and ¹³C-NMR experiments. It contained hydroxyl and carbonyl groups due to the IR absorption bands at 3441 and 1686 cm⁻¹, respectively. The ¹H-NMR data of **1** showed four singlet methyl groups (δ_{H} 0.85, 1.03, 1.16, and 1.59), one doublet methyl group [δ_{H} 1.08 ($J = 6.5$ Hz)], two oxygenated methine protons [δ_{H} 4.17 (br s, H-3), 4.34 (br d, $J = 11.5$ Hz, H-2)], two oxygenated methylene protons [δ_{H} 3.75 (d, $J = 11.0$ Hz, H_a-23), 3.90 (d, $J = 11.0$ Hz, H_b-23)], two olefinic protons [δ_{H} 5.44 (dd, $J = 1.5, 9.5$ Hz, H-11), 6.30 (dd, $J = 1.5, 9.5$ Hz, H-12)] at low

field region (Table 1). The ^{13}C -NMR and DEPT data showed four oxygenated carbons (δ_{C} 66.6, 71.7, 75.9, and 79.5), a disubstituted double bond (δ_{C} 119.6 and 142.1), and an acid carbonyl functional group (δ_{C} 181.2). On account of the molecular formula $\text{C}_{30}\text{H}_{46}\text{O}_6$, the index of hydrogen deficiency (IHD) of **1** was eight including one acid carbonyl and one olefinic functionalities. Thus, the number of rings of **1** should be six. In HSQC and HMBC of **1** showed that two cyclopropyl protons at δ_{H} 1.52 and 2.35 had long-range correlations with one of the olefinic carbons (δ_{C} 142.1, C-12), three quaternary carbons [δ_{C} 28.2 (C-13), 33.6 (C-14), and 34.5 (C-8)], one tertiary carbon (δ_{C} 47.8, C-18), and one secondary carbon (δ_{C} 22.8, C-15). All these data suggested that compound **1** was a 13,27-cycloursane-type triterpene acid with four hydroxyl groups at C-2, -3, -19, and -23, one acid group at C-17, and one double bond at C-11 and -12. The NOESY spectrum of **1** exhibited mutual correlations between H-2, H₃-24, and H₃-25, confirming H-2 to be β -oriented. H-3 was also deduced to be β -oriented to fit the small axial-equatorial coupling constant between H-2 and H-3. Further analysis of all the 2D NMR data allowed the complete assignment of ^1H - and ^{13}C -NMR spectra of **1**, and these results are listed in Table 1. Accordingly, compound **1** was established as 2 α ,3 α ,19 α ,23-tetrahydroxy-13,27-cyclours-11-en-28-oic acid. Although ursane triterpenes were commonly found in higher plants, 13,27-cycloursane-type triterpenes such as **1** are rare. To our knowledge, the other three analogues of the 13,27-cycloursane triterpene were isolated from *Phyllanthus engleri*,¹¹ and *Ficus microcarpa*.¹²

Compounds **1–7** were evaluated for their cytotoxic activities against two cell lines which were named human colorectal carcinoma HT29 and human breast carcinoma MCF-7. After 72 h of treatment, the relative polar compounds **1** and **2**, both bearing four hydroxyl groups at their C-2, -3, -19, -23 and one acid functionality at C-17, with IC_{50} values higher than 40 μM seemed to be less cytotoxic than compounds **3–7** toward two cell lines (Table 2). Compounds **3–7** exhibited IC_{50} values ranging from 5.8 ± 1.4 to 32.4 ± 3.8 μM , and among them, **4** and **7** were the most toxic. Concerning the structure and activity relationships, pentacyclic triterpenes bearing a carboxylic acid functionality at their C-17 were found to exhibit potent cytotoxicity in literatures.¹³ However, **1** and **2** exerted comparably low cytotoxic effect toward HT29 and MCF-7 in our results. The reason why **1** and **2** exhibited less potency toward these two cell lines remained to be studied. In addition, it was shown that pentacyclic triterpenes with an (*E*)- or (*Z*)-coumaroyl functionality at C-3 or -23 would display significant cytotoxic effects.¹⁴ This was also observed in **7** which possessed an (*E*)-coumaroyl at its C-3, and exhibited an IC_{50} value similar to the positive control, the clinically used anticancer drug etoposide

(VP-16, $IC_{50} = 3.6 \pm 1.7 \sim 8.2 \pm 3.8 \mu\text{M}$).

EXPERIMENTAL SECTION

General Methods

Optical rotation was measured on a Jasco P-1020 polarimeter (Tokyo, Japan). IR spectra were recorded on a Thermo Mattson IR300 spectrometer (California, USA). ^1H , ^{13}C and 2D NMR spectra were acquired on a Bruker DMX-500SB spectrometer. LR/HRFABMS and EIMS were obtained on a Finnigan/Thermo Quest MAT-95XL spectrometer (Bremen, Germany). HPLC was performed on a Hitachi L-7000 liquid chromatograph with a Bischoff RI detector (Leonberg, Germany).

Plant Material

The leaves of *P. duclitan* were collected from National Museum of Natural Science at Taichung in December, 2003, and were identified by Dr. Chen Chang, an assistant researcher in Department of Botany, National Museum of Natural Science. A voucher specimen (No. 12292003) has been deposited at the Graduate Institute of Pharmacognosy Science, Taipei Medical University, Taipei, Taiwan.

Extraction and Isolation

Fresh leaves (3.0 kg) of *P. duclitan* were extracted three times with 10 L MeOH at room temperature for two weeks. The methanolic extract was concentrated in vacuum to give a black residue (200 g), which was re-dissolved in 85% aqueous methanol and then partitioned with *n*-hexane to generate two fractions: the aqueous methanol soluble fraction and *n*-hexane soluble fraction. Subsequently the *n*-hexane fraction was vacuum-evaporated to dryness (50 g), which was pre-adsorbed with 75 g of silica gel, then loaded into a Si-open column (8 × 26 cm) with mixtures of *n*-hexane and EtOAc as eluents in a step-wise elution mode. Every 300 mL of eluent was collected as one fraction and each was analyzed by thin layer chromatography using plates of Silica gel 60, F₂₅₄, 0.2 mm thickness (Merck, Germany), and a solution of EtOAc/*n*-hexane (2:1) for development. Vanillin-sulfuric acid charring to form purple or blue spots was used to detect the triterpene acids. Totally 78 fractions were collected, and all the fractions were combined into nine major portions according to the TLC results. Portion VI (#fr.41 ~ 50) eluted by EtOAc/*n*-hexane (3:7) was further purified by repetitive HPLC on a Hypersil ODS semi-preparative column (10 × 250 mm, Thermo Electron Corp., Bellefonte, USA) with MeCN/H₂O (85:15) containing 0.1% trifluoroacetic acid (TFA) as eluent to afford **4** (68 mg) and **5** (14 mg). Portion VII (#fr.51 ~ 58) eluted by EtOAc/hexane (1:1) was further purified by HPLC using the same column with MeCN/H₂O (80:20) containing 0.1% TFA as eluent to afford **3** (37 mg) and **7** (8 mg). Portion VIII (#fr.59 ~ 70) eluted by EtOAc was further purified using the same chromatograph with MeCN/H₂O (65:35) containing 0.1% TFA as eluent to afford **6** (5 mg), and with MeCN/H₂O (45:55)

containing 0.1% TFA as eluent to afford **1** (16 mg) and **2** (14 mg).

Cell Culture

Human colorectal carcinoma HT29 cells and breast carcinoma MCF-7 cells were maintained in RPMI-1640 medium supplied with 5% fetal bovine serum, 100 units/mL penicillin and 100 µg/mL streptomycin.

Growth Inhibition Assay

Cells in logarithmic growth phase were cultured at a density of 1×10^4 cells/mL/well in a 24-well plate. The cells were exposed to various concentrations of the test drugs for 3 days. At the end of the incubation period, cells were fixed and stained with 50% ethanol containing 0.5% methylene blue for 30 min. The plates were washed five times with water and allowed to air dry. The resulting colored residue was dissolved in 1% *N*-lauroyl-sarcosine, and optical density was read at 570 nm using a Bio-Rad microplate reader (Model 2550). Each point represents the average of at least two independent experiments run in triplicate.

2 α ,3 α ,19 α ,23-Tetrahydroxy-13,27-cyclours-11-en-28-oic acid (**1**)

Amorphous white powder; $[\alpha]_D^{25} -6.5^\circ$ (*c* 1.0, MeOH); IR ν_{\max} (KBr) 3441, 2929, 1686 cm^{-1} ; ^1H - and ^{13}C -NMR data see Table 1; FABMS m/z 503 $[\text{M} + \text{H}]^+$; HRFABMS m/z 503.3376 $[\text{M} + \text{H}]^+$ (Calcd. for $\text{C}_{30}\text{H}_{47}\text{O}_6$, 503.3373).

ACKNOWLEDGEMENTS

This study was supported by grants to Dr. T.-H. Lee of the National Science Council (NSC 93-2113-M-038-003). We are grateful to Ms. Shwu-Huey Wang and Ms. Shou-Ling Huang for the NMR data acquisition in the Instrumentation Center of Taipei Medical University and Instrumentation Center of the College of Science, National Taiwan University, respectively.

REFERENCES

1. Yang, T. Y. A. Sapotaceae. In: *Flora of Taiwan*; Editorial committee of the Flora of Taiwan, Department of Botany, National Taiwan University: Taipei, Taiwan, 1998; Vol. 4; p 85.
2. Wandji, J.; Tillequin, F.; Mulholland, D. A.; Shirr, J. C.; Tsabang, N.; Seguin, E.; Verite, P.; Libot, F.; Fomum, Z. T. *Phytochemistry* **2003**, *64*, 845.
3. Taniguchi, S.; Imayoshi, Y.; Kobayashi, E.; Takamatsu, Y.; Ito, H.; Hatano, T.; Sakagami, H.; Tokuda, H.; Nishino, H.; Sugita, D.; Shimura, S.; Yoshida, T. *Phytochemistry* **2002**, *59*, 315.
4. Gao, H.; Wu, L.; Kuroyanagi, M.; Harada, K.; Kawahara, N.; Nakane, T.; Umehara, K.; Hirasawa, A.; Nakamura, Y. *Chem. Pharm. Bull.* **2003**, *51*, 1318.
5. Taketa, A. T. C.; Breitmaier, E.; Schenkel, E. P. *J. Braz. Chem. Soc.* **2004**, *15*, 205.
6. Deng, J.-Z.; Starck, S. R.; Hecht, S. M. *J. Nat. Prod.* **1999**, *62*, 1624.
7. Cheng, D.-L.; Cao, X.-P. *Phytochemistry* **1992**, *31*, 1317.

8. Kim, N.-C.; Desjardins, A. E.; Wu, C. D.; Kinghorn, A. D. *J. Nat. Prod.* **1999**, *62*, 1379.
9. Numata, A.; Yang, P.; Takahashi, C.; Fujiki, R.; Nabae, M.; Fujita, E. *Chem. Pharm. Bull.* **1989**, *37*, 648.
10. Häberlein, H.; Tschiersch, K.-P. *Phytochemistry* **1994**, *35*, 765.
11. Alberman, K. B.; Kipping, F. B. *J. Chem. Soc.* **1951**, 2296.
12. Chiang, Y.-M.; Su, J.-K.; Liu, Y.-H.; Kuo, Y.-H. *Chem. Pharm. Bull.* **2001**, *49*, 581.
13. Chiang, Y.-M.; Chang, J.-Y.; Kuo, C.-C.; Chang, C.-Y.; Kuo, Y.-H. *Phytochemistry* **2005**, *66*, 495.
14. Chang, C.-I.; Kuo, C.-C.; Chang, J.-Y.; Kuo, Y.-H. *J. Nat. Prod.* **2004**, *67*, 91.

Table 1. ¹³C- and ¹H-NMR spectral data of compound **1** (500 MHz, pyridine-*d*₅).

position	δ_C (ppm)	mult. ^a	δ_H mult. (J/Hz) ^b	position	δ_C (ppm)	mult. ^a	δ_H mult. (J/Hz) ^b
1	43.4	t	1.80 2.19	16	26.9	t	2.02 m 2.72 m
2	66.6	d	4.34 br d (11.5)	17	47.5	s	
3	79.5	d	4.17 br s	18	47.8	d	2.85 s
4	42.4	s		19	75.9	s	
5	44.1	d	2.15	20	42.9	d	1.48
6	18.9	t	2.00 1.70	21	27.4	t	1.40 1.96
7	37.7	t	1.49 1.92	22	38.0	t	2.07 2.18
8	34.5	s		23	71.7	t	3.75 d (11.0)
9	53.8	d	2.17				3.90 d (11.0)
10	38.3	s		24	17.6	q	0.85 s
11	119.6	d	5.44 dd (1.5, 9.5)	25	19.4	q	1.03 s
12	142.1	d	6.30 dd (1.5, 9.5)	26	16.9	q	1.16 s
13	28.2	s		27	16.5	t	1.52 d (4.6)
14	33.6	s					2.35 d (4.6)
15	22.8	t	1.75 m 2.54 m	28	181.2	s	
				29	27.4	q	1.59 s
				30	16.3	q	1.08 d (6.5)

^a Multiplicities were obtained from DEPT experiments.

^b Signals without multiplicity were picked up from COSY or HMQC spectra.

Table 2. IC₅₀ values of compounds 1–7 against human HT29 and MCF-7 cancer cell lines.

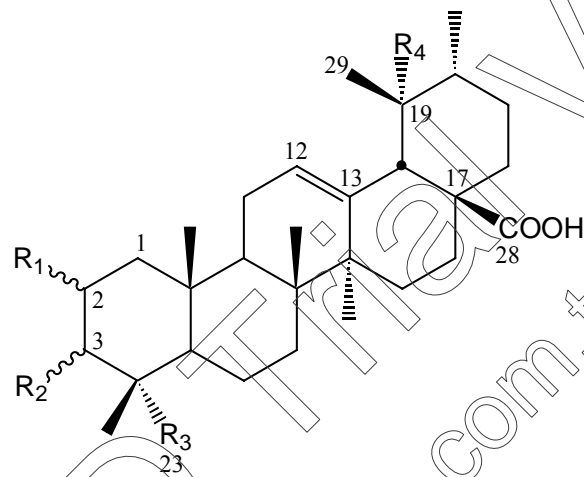
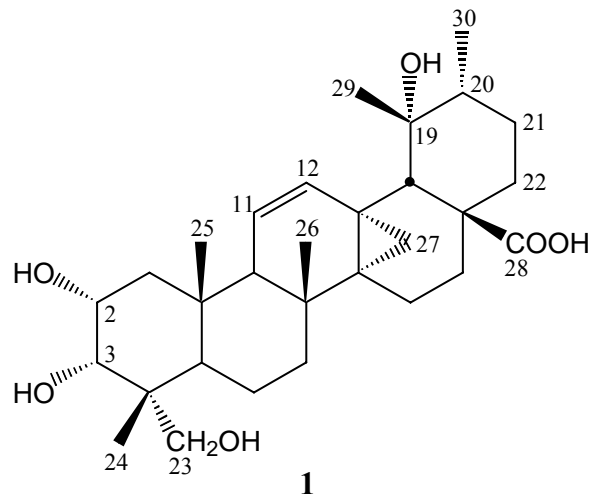
compound	IC ₅₀ (μM) ^a	
	HT29 ^b	MCF-7 ^c
1	> 40	> 40
2	> 40	> 40
3	6.9 ± 2.3	15.1 ± 2.1
4	5.8 ± 1.4	6.3 ± 0.5
5	24.1 ± 4.2	32.4 ± 3.8
6	21.8 ± 3.6	9.5 ± 2.5
7	6.5 ± 1.9	6.3 ± 1.7
VP-16 ^d	3.6 ± 1.7	8.2 ± 3.8

^a Cells were treated with various concentrations of tested compounds for 3 days. Cell growth was determined by methylene blue dye assay. The IC₅₀ value resulting from 50% inhibition of cell growth was calculated. Each value represents the mean of three independent experiments.

^b HT29 as human colorectal carcinoma cell line.

^c MCF-7 as human breast carcinoma cell line.

^d VP-16, a chemotherapeutic drug, as reference compound in this study.



	R ₁	R ₂	R ₃	R ₄
2	α-OH	α-OH	CH ₂ OH	OH
3	α-OH	β-OH	CH ₃	H
4	H	β-OH	CH ₃	H
5	H	β-OH	CH ₃	OH
6	H	β-OH	CH ₂ OH	OH
7	α-OH	β- <i>O-trans</i> -coumaroyl	CH ₃	H

計畫成果自評：

本研究計畫預期能由蘭嶼山欖葉部找出能毒殺乳癌細胞株MCF-7 的活性物質，而根據研究成果顯現蘭嶼山欖葉部的低極性部分，確實含有多種三萜酸類 (triterpene acid)，其中部分化合物對於乳癌細胞具有不錯的毒殺效果 ($IC_{50} = 6.3 \pm 0.5$)，活性強度與臨床用藥VP-16 ($IC_{50} = 8.2 \pm 3.8$) 近似，此外，其中一個三萜酸含環丙基的骨架，為三萜類中較少見的新化物。經評估此成果與本計畫預期相符，同時具有應用價值，所有內容業經投稿於Journal of the Chinese Chemical Society (SCI)，並已獲得接受發表，預計於明年初 (2006) 刊出。

PDFCMD Trial Version
www.zeon.com.tw