ƒ ‹ F | Œfia‹ ·'e›ß• |–M^D‹ ª¤s›p e ƒ¤"G†ŁSi

⁄ j¥ „«⁄"¯ƒ¥H⁄ ¸¥ ⁄Ufl " † ›M¥Hokadaic acid¤°¿Efl ‰Ł⁄ , ⁄Sx ª¤s

›p e^ ®¡G SO«‹ ›p e

›p e‰ ¸„¡GNSC91-2314-B-038-046-

ɹf · ¶¡¡G91 ƒ~08⁄°01⁄ Ø 93 ƒ~07⁄°31⁄Ø

ɹf † ƒ ¡G» O ¥´ ·⁄j ·, › ·‹

›p e¥ D ø⁄ĦG¶ z†[

›p e »P⁄H›ß¡G† fl « ‹ ª¤s¥˝

†ŁSi^ «‹¡G" °††ŁSi

† B † ƶŁƒG¡¥»›p e¥ i ⁄ ‰ ¶‹ d,

⁄⁄ ¥` Œ 93 ƒ~12⁄°20⁄Ø

ƒ ‹ F | Œfia‹ ·'e›ß•|, §U–M^D‹ ª¤s›p ef¤"G†ł§i

⁄j¥ „«⁄"¯ƒ¥H⁄ ¸¥ ⁄Ufl " † ›M¥Hokadaic acid

¤°¿Efl ‰Ł⁄ , ⁄§‹ ª¤s

›p„"‰s, „¡GNSC 91-2314-B-038-046

ɪf ·`¶¡¡G91 ƒ~08 ⁄°01 ⁄Øf 93 ƒ~07 ⁄°31 ⁄Ø

¥D«ø⁄H¡G¶ z† [˘—— ´

›p„" »P⁄H›ß¡G†fk« ‹ ª¤s¥"

ɪf "c⁄ ¸† ƒ ƒW" ¡G¥x¥_´ ·⁄j ·, › ·‹

⁄@¡B⁄⁄ "K›n

fi L„¥h™"‹ ª¤s– o "¡Fl ' ⁄ "ƒ¥Ł¥ ⁄U⁄§fl " † ´ · ¡ A g¥ (ƒp",»X catecholamines) " " ¤ ° ¿ E ¡ A ¥ i ¥ H « P ¶ i fl " † › M ¶ i ƒflcAMP , ¡C¤ §@¥¸ T fi ¶·¡A‹ ¸˘cyclic AMP-dependent protein kinase (PKA)¡ ¶iƒ «P¤˘† M‰Ł⁄"" hormone-sensitive lipase (HSL) ` ß˜⁄˘ƒ ‹ ¸˘¡A¤^¥ † ›M‰Ł †ffl " " w " ‹«P¤˘fl ‰ Ł ⁄ , ƒ¡G¥~¡ ŒHphosphatase "⁄"§ ¤ flokadaic acid ¤ ° ¿ E fl " † ›M⁄"¡A⁄† ¥›M ‰Ł• | o ¥ "«ß˜⁄˘"⁄† ¶H¡Aƒ P fi ⁄ ł |fł ‰ Ł ⁄,ƒ ¸©ƒ„^ "«" " ß¥¤¸^⁄ £ ł...W ¥[HSL "⁄‹ ¡'°¡Aƒ ‹ O ®¶HSL ´ †ffl " " w " ›¡Cƒ]ƒ„fl " " w " ›⁄† ¥ ‰Łƒłfl ‰Ł ⁄ , ⁄Ł›–§ Œ " ŧ«\⁄fª ƒ ¡C" æ ƒ¤ o ł⁄⁄"ffl " † ›⁄M⁄"HSL ‹ ¡'°, ß ¥⁄fU " † ›M "⁄"HSL ‹ ¡'° "¡ F ¥ ¥ † ›M⁄"fl " " w " ›⁄† ¥ ‰Ł¥ ⁄£" ‹ ƒP¡C' ¥H¥ » „ Œ ¥-˜⁄j¥ „«⁄"¯ƒ¥H⁄ ¸¥ ⁄Ufl " † ‹›M §fi¡˜Afbisopreterenol ¥H⁄ ¸okadaic acid ¤ ¿E« «Æ¡A ;œ ¤^⁄æ,ßfl ‰ Ł «ŧÆ" " fl"»˜˜ æ¶pC ф{ H ¿«⁄ W ¡A· ´ fl " »˜"⁄"§ t ¶‰„ŒE..W ¥[¡Cƒ §Q¥¸SDS-PAGE „ ¢a⁄Ł"k⁄ "Rfl " " w " ›⁄† ¥ ‰ Ł"¯ ⁄˘¡A⁄] o†ffb isopreterenol ¥H⁄ ¸okadaic acid ¤ ¿E«Æ¡ A fl"⁄" w " ›⁄† ¥ ‰ Ł"" ^»§P ¶q» PL^ "«¤° ¿ Ł"„ •†⁄æ,ß⁄]ƒ⁄ ŧP"⁄¯ ⁄˘¡Cƒ ' ‹ O†£¥´¯ ⁄˘" ⁄Of " † ¥ ‰Ł« h «§¶i⁄ @B"" „ Œ ⁄ "R¡C

^ `ƒ:okadaic acid, fl ‰ Ł „fl " † ›M

Abstract

Several studies have shown that lipolytic hormones such as catecholamines stimulate lipolysis in visceral and subcutaneous fat cells. The mechanisms are mediated by increasing in intracellular cAMP concentrations and then activating cAMP-dependent protein kinase (PKA) which induce phosphorylation and activation of hormone-sensitive lipase (HSL). The activated HSL translocates from cytosol to the surface of intracellular lipid droplets to promote lipolysis. Okadaic acid, an inhibitor of protein phosphatases type-1 and type-2A, is also found to stimulate lipolysis in adipocytes. It does not increase

activity of HSL but induces translocation of HSL to the lipid droplets, thus the surface proteins located on lipid droplets may be closely related to this lipolytic action. Recently, visceral fat cells are found to contain higher HSL activity than the subcutaneous fat cells. There is also considerable difference in protein pattern between visceral and subcutaneous lipid droplet preparations. In this study, we found the increase of free fatty acid during isoproterenol- or okadiac acid-stimulated lipolysis by spectrophotometry. The protein pattern and protein content of lipid droplet fractions following isoproterenol- or okadiac acid-stimulated lipolysis were different from that of control by SDS-PAGE. The changed proteins could be identified in advanced studies.

Keywords¡Gokadaic acid, lipolysis, fat cells

⁄G¡B†‡Si⁄″fie

1. «e¤¥»P‹ ª¤S¥ ″″

fl ″ † ´ · ¥ D › n ¤ ƒ†(«fOp¯@f† '…ƒh″c†¸ȳ)₣ ₣¸«¥O•x″″¥ \ fl Ą.ffl ¥₣Ł ⁄ , (lipolysis)† ¥˝″″fl ″ »˜(free fatty acid)⁄] ‹ O ¤¯Øn″«fl ¶ q ¤ • ‰ S⁄f@¡″C¤Ø • ƒ ₧›⁄ØSG″″ƒ ,m¥ Dn⁄ ‹ ¤ ⁄j¡^A⁄ @ ^ ‹ O ƒ'† ¥.h»Sfl ″ † ″‹ subcutaneous type¡A¥t⁄ @ «h‹ O ƒ'⁄″¯ƒ' ₱ ″″fl ″ † ″‹ visceral type (Leibel et al., 1989)¡G¥ ' visceral type ″″fl ″ † ¤ fl ‰ Ł ⁄,† ¥˝″″fl ″ »˜⁄jf ₧g¥ ¤ x ″ ØRfl (hepatic portal vein)¶ ŧJ¤x¯ƒ¥ ₣¸· ` ¤t⁄†″¡Aƒ ƒ„ƒ† · ″ – ‰isceral type ″″fl ‰Ł ¥ Ñ´»P¿ } Sfl₣ ₣¸ °fl w˘ ¥fl fl₣ƒ†,ß⁄j″″^ þ'(Bjorntorp, 1985)¡C

L‰ ‹ Ovisceral type ' ¿‹ Osubcutaneous type ″″fl ″ † ›M¸Afb¤ ¤ †″',»X˜ ₰¤° ¿Efl ‰ Ł ⁄,fi ¡A¤ T fi S ¶ · ,‹ Ği⏐¥ † ″»₰P† › ₥⁄″W¤ „ † ₣«₣A¤ ˘o † ›M⁄″″cyclic AMP (cAMP)¿ ₰⁄W ¡A‹ ¡⁄˘† ›M‰Ł⁄″″cyclic AMP-dependent protein kinase (PKA)¡A¶if «P¤ ˘ † ›M″› ₮dfl ‰ Ł ⁄,″″» fl hormone-sensitive lipase (HSL)` O»˜⁄¥ ₣¸ ¡⁄˘(Olsson et al.,1987)¡ Ą ‹ ¡⁄˘″″HSL • ⏐¥ † ›M‰Ł⁄⁄ † ¤ fl ″ ″w″″ ›–¡A¶i⁄@¤ BNfl ‰ Ł ⁄,ƒ ¤ ¥˝(glycerol)»Pfl ″ »˜(free fatty acid)˜ ¥X¡Aƒ ƒ„fl ‰ Ł ⁄,″″„ L{⁄⁄HSL ″″^ O»˜⁄˘»P‹ ¡⁄˘¤ ƒ†›«›n″″¥ Ñ (Yeaman et al., 1994)¡C

″ æ~¤ †\ƒh″″‹ ª¤s† ~ ofk₣fl ″ † ›M⁄″″″fl ″ ″ w ″ ›ƒ††\ƒh″″† ₩ ‰Łfp perilipins¡Badipose differentiation-related protein (ADRP)»P capsular protein ¥¡A‹ ¤ ƒ† ‰ –⁄fl ‰Ł¥ Ñ´″″¥ Ñ (Londos, 1999)¡CPerilipins ‹ O† ƒ†Q ф{″″fl ″ ″ w ″ ›⁄– ₩ ‰ ₤ĄA ƒsƒ ₰ fl ″ † ›M»P' TJ^ ⁄ ″c† ›M^″″fl ″ ″ w ″ ›(Greenberg et al., 1991¡BServetnick et al., 1995)¡C• † › M ¤ ¤ † ″,•»₰¤Q†₫KEA¡` O»˜⁄˘¡A†Q»{‹ ¤ ƒ† SUHSL ″ ' fl ″ ″ w ″ ›″″¥ Ñ (Londos et al., 1995)¡CAdipose differentiation-related protein (ADRP)«h‹ O• ¶Mƒsƒ ₰ †\ƒh⁄ £ ₣″^ ″″† ›M⁄⁄¡Afp adipocytes, steroidogenic Leydig cells, fibroblasts »P hepatoma cells¡C⁄ ₩ ƒ„† ₩ ‰Ł¥i¥Hk«P¶i† ›MS l ƒ ‹ ₤fl ″ »˜ (long-chain fatty acid)¡AS ¸ƒ¤fl ″ ″w ¿n¡A¤ ¥]† ƒb″ S ¸ƒ¤″″fl ″ ″w' ₱ ¥[¥Hk‹ O@ (Brasaemle et al., 1997)¡Cƒ †Q¯n† ₩ ‰Ł(capsular protein)«h‹ OS › „ Œ « ₫'{¡Afs ƒ ₰ ' TJ^ ⁄ ″c† ›M^″″fl ″ ″ w ″ ›¡–A• † › M ¤ ¤ † ″,›₰¤¸ª ¿₰Łfi‰Ł•⏐–qfl ″ ″ w ″ ›⊢ ,¤¡A· ´ ƒb† ›M‰Ł⁄⁄¡A– ·∞»Pfl ‰ Ł ⁄,ƒ†^ ¡C(Wang and Fong, 1995¡EFong et al., 1996)¡C† ″æS › ″″‹ ª¤s ф{ƒbfl ″ † ›M»P ˙⁄W, ¢¥ ‰Ł† ›M^″″fl ″ ″ w ″ ›ƒ†¤'

¥ L « ‹ ƒ °(beta-actin)" " ƒ(Dong et al., 2001)¡ C ƒ] ‹ ƒ ‡ " ƒ·¿,g¥‡"
ƒ h fi ‰ (anti-actin antibodies)' ¸¥H^ "«cytochalasins ‡ B † z · ⁄ W , ¢ ¥‰Ł† °›M⁄z´Z
‡ ¥ « Æ ¡ Ao‡ACTH ' ⁄ o"fl ‰Ł⁄ , †{¶ H ⁄• |o¥ ˝ ¡ F ƒ‹‰Ł⁄ , '˜"¥X‡T
J(cholesterol)« o L " k „ Be¯ƒØ†⁄‰Xƒ¤ · ⁄ W ¢¥ «(cortisols)¡ C]ƒ „ – ·œ¥‰‹
ƒ ‡J¥ i fl»P ‰Łƒb› M⁄"""„ Beƒ‡^ (Mrotek and Hall, 1977; Hall et al., 1981)¡C
¥ ƒ ¥i¤£¡Afbfl ‰Ł⁄ , "„ L{⁄⁄ £P"fl " " W ‡›¥ ‰Ł‰ C " t ‡‡›» «n"¤ ƒ ¡C
Okadaic acid ‹ O⁄ @ serine/threonine phosphatase typ-1 »Ptype-2A "‰ ¤ £A¥H
ƒ„^ "«¤ ¿E† ›Mi • | ›P† ›M⁄"""‡\fh‡ ¥ ‰Ł¥X†{ " « C»"‰"†{¶H (Bialojan et
al., 1988; Haystead et al., 1989)¡C⁄ W okadaic acid ¤ ¿Efl " † M¡¥Ai¥H« ¶£ifl ‰Ł⁄ ,
"‰ ¶¿¡Af ¥ £ fl " " W "Y–perilipins ⁄]ƒ‡` C ›⁄"""†{¶H(Clifford et al., 1998)¡C
¥H⁄"¯ffl " † ´·(epididymal fat pad)‹ ‰ fi˘¡A¶i⁄@¤B⁄ "Rƒ„^ "«"‰ ¶¿ ´ fi ¡ A o†{
okadaic acid "‰¤ ¿E• |¤ ˘oHSL ` C ›⁄¡Af ‹O¤^⁄£• |¤˘† ›M⁄"total HSL "‰‹ ¡ ' …W¶¥A[
ƒ ‹O« ¶£i` C ›⁄""HSL ´ † ƒ fl " " W (Morimoto et al., 2000)¡Cƒ„¥~¡A– N H⁄fuX
ƒ¤"fl " "w(pure lipid droplets)»P` C ›⁄""HSL ƒ@ƒP ¤|¡A¤^⁄£• ¥X†{fl ‰Ł⁄ , "‰†{
¶ £F⁄˘ƒ ‹O–q† ›M⁄⁄ ´ ¥X"fl " "w(crude lipid droplets)»P` C ›⁄""HSL ƒ@ƒP
¤|¡A⁄~• | o¥ fl ‰Ł⁄ , ¡ C ·"› ƒ]ƒ „ ‰ ¡Afl " † ›M"fl ‰Ł⁄ , S ¶¿¡A £ P»›n« ¡
⁄˘"""HSL ⁄S¥~¡A¥D›n¤ Mwƒ]fl ‹Ofbfl " "w"" "c¡A¥ SYfl " " W "Y‡ ¥ ‰ŁS Ct‰
––fl ‰Ł¥ Ń´"›« › ¤⁄ƒ(Morimoto et al., 2000)¡Cƒ ‹O¥Hokadaic acid ¤ ¿E«Æfl " "
›–‡ ¥ ‰Ł"‰ ¶q»P" ^ ƒ‡ L¯ ⁄˘¡A«o' ¥..ƒ‡⁄ m̃‡Ł ¡C¥t⁄@⁄Ł›–¡A⁄wf‡‹ ª ¤ s ‡Si«
¥X¥H« ƒP¿@« "¥¿ · W,f¢ (norepinephrine)¤ ° E£fi¡A⁄"¯ffl " † ›M⁄› ¢dfl ‰Ł⁄ ,
""HSL ‹ ¦°¡A⁄æ¥ ⁄Ufl " † ›M⁄"HSL ‹ ¦°‹ "¡Ff ¥B† ›M⁄"fl " " W ‡›¥ ‰Ł
"" ^ »P‰ ¶q⁄]⁄£" ‹ ƒP(Morimoto et al., 1997)¡C‡ "æ‡\f h " ¥H»«"3T3-L1 fl
" « ¤›† ›M« ‰ fi˘¡Afb^ "«isoproterenol "‰ ¶¿U¤ ¿Efl ‰Ł⁄ , ¡A⁄æ,ß¤ ¿E«e«Æfl
" " W ‡›¥ ‰Ł"¯ ⁄˘¡C ¢{⁄£‰ ¤ ¿E«e«Æfl " " W ‹›–ƒ‡perelipin, S3-12,
vimentin ¥H⁄¸TIP47 ¥ "c'°‡ ¥ ‰Ł¡C‹ ⁄˘ƒa¡Aadipophilin, caveolin-1 ¥H⁄¸tubulin
¥ "c'°‡ ¥ ‰Ł¥u¥X†{ƒb¤ ¿E«Æ"fl " " W ¡C¥t¥~` ƒ‡‡\fh»Pfl ‰Ł¥ Ń‹ ^ "‰†J
¥ ‰ ŁAfb^ "«¤ ¿E«e«Æ]‡£ƒ‡⁄ £P"¯ ⁄˘(Brasaemle et al., 2004)¡C
' ¥H¥ » „ Œ ¯ ›p› e§¥H⁄j¥ „« ‹ ¤s„ ¶H¡A⁄ § ©¤¥X⁄"¯ƒ(epididymal fat pads)
»P¥ ⁄U(femoral subcutaneous fat tissues) ¥‡¡ƒ "fl " † ´·¡A⁄ ´ ¥ ¤ " † M¡A„'
okadaic acid ¤ ¿E¡A« ¶£ifl " † ›M⁄"""HSL »Pfl " " W ‡›¥ ‰Ł` C ›⁄¡A¤^⁄ §O ¸
·œfl " »˜"‰˜ 'æ¶qƒ‡ L¯ª fi ŧS¡Cƒ ¥Bƒb¤ ¿E«e«Æ⁄ ´ ¥X† ›M⁄"""fl " "w¡A¥H„ Ča
"k(SDS-PAGE)⁄ "R" ›–‡ ¥ ‰Ł"‰ ¶q»P" ^ ‹O§ § ¯ ¡C§ ˘ –´¥ ƒ„„ Œ ¶i⁄@¤B⁄F,
ƒ ' ⁄"¯ƒ»P¥ ⁄U"fl " † M¡A fl ‰Ł¥ Ń´"" ¤ »P" ›–‡ ¥ ‰Łƒ‡ƒ ⁄ £P¡A¥H ‹ –N
¤ ‡ fQ ‰Ł‹ ^ fl ¤lf"" ´ƒ¡C

2. ‹ ª¤S⁄Ł"k

¡]1¡^„Œ¯ °"«

¤œ250-300 § J › « §¶M¡star rat¡A „ } i ƒ b ‡ q › • ¤ } ƒ n " " „⁄U¡A¤^ „⁄'¥¿–
¥Hsodium pentobarbital (50 mg/kg body weight), g , ¡ ˜"›fl¥G‡ ´K¡A⁄ § O ¤ œX⁄"¯ƒ
"fl " † (epididymal fat pad)¥H⁄ ¥ ⁄Ufl " † (femoral subcutaneous fat tissue)¡C

¡]2¡^fl " † ›M"/ ´

fi 1964 ƒ~Rodbell "/ Ł " ¶ifƒ " † ›M"/ ´ ¡ C S @ ƒ p / ŲG"N/j¥ „« ¯ffl " † ´·(epididymal fat pads)¥ H ¿¥/Ufl " † ´·(femoral fat tissues)¡A/ S O m' normal saline (0.85 %/S NaCl)/ / … ‹ «Æ¤Œ¥Xfl «ÆNfl " † ´·, m albumin-bicarbonate buffer(St 4% BSA, 3.3 mg/ml collagenase type II /¸ 3£gmoles glucose, pH 7.4)¡A¤ • w/v = 1:1 / æ ¡C¤ ^ ¥ /p–Nfl " † ´· ¯ ƒ ¤p¶Afb 37°C / / ˇ+1 /pfi ¡Cƒ „ fi †‰l´·"/‰ ƒ › ¯ "ß¡¥Afl," † ›M• ƒ fl " † ´·/ / ´ ¥X¤ ¡A ¤ ˇ – o ‰ w ‰ †G¡TC¥HV¿H–Nf¤¥ ƒ ¶ " ¥…/ , /Sfl " † ´·¶ ¤ Œ¥X¥ Æ «Æ¡A¥H400 g/S´ +t´ / 1 / ˜`¡A« ¥i/ ´ ¥X/ Wh/Sfl " † ›M¡A/U..h(¥]St/W¥ † ´·,/"¥ † ›M, ¥ ¤‰† ›M ¥)«h¥ Æ¡ €N/ Wh/ ´ ¥X/Sfl " † ›M¥H37°C "/‰ w ‰†.G ‹«¡Æƒ A¥H400g ´ +t´ / 1 / ˜`¡Afl„†M‹ ~¤ BJ›«‰˜3 ƒ,«Æ¡Afl ¶ / Wh¥ ƒ "/fl " † ›M¡A¶ ƒ „ Œ ¡C

¡]3¡^Okadaic acid "/¤°¿E

– N » s + ˇ S · /S fl " † › M ¡ A • n / ^ « Æ ¥ ¶ q Buffer, A(St 2.5% / /¡A¥t¥~¥[/J BSA, 135 mM NaCl, 5 mM KCl and 1 mM $MgCl_2$, 25 mM TES, pH 7.4)¡ A ƒ Æ[/J okadaic acid¡ F ˇ okadaic acid + † @«0¡B3¡E10¡B30 µM¡A' 37°C / / ˃ +2 / p fi¡C ¥H 400g / S ´ t ´¡Afl ‹ ¶ / W … h / S fl " † M¶¡Aƒ fl …" h"w/»S/+ Œh¶i ƒ fl " »˜(free fatty acid)/S »œ¡C

¡]4¡^ ¸·œfl " »˜/S·œ'w/Ł"k

¥ » „ Œ ¯–˜1986 ƒ~ Okuda ¥ " /"Ł"k¡ A»·œfl " »˜" " ˜ ' æ ¶ q Ç¥ –N/W ›z okadaic acid ¤ ¿Efl " † › M Æƒ‹¶• »G(‹ øœ 250£gl)»P 3ml chloroform-heptane 1:1(St 2% methanol) † ¥X¡A¤ ^ ¯ " 'S¦0 ˆmin¡Afl Æ H2,000 g ´ + t 5 / ˜`¡A¥H ' Sl"/Łƒ¡– NW.. h • »G.. h S l ¥¥¡E€N1 ml "Copper reagent ¥ [ƒ/.Lhf + • »G …h¡AfAƒ,¯ ' 10 min¡A¤ˆ¥H2,000g ´ + t 10 min¡C¤œ/W..h" ‰Ł‹ ø0.5 ml »P0.5 ml chloroform (/"St 0.1% bathocuproine »P 0.05% butylated hydroxyanisole)† V ƒ S ¡¡Af «Æ¥H" Ł‡480nm ·œ¶qfl " »˜/S˜ ' ¶q¡C

¡]5¡^† ›M/"fl " "w/S/ ´

– N ƒ ‹ ¶ " " £ffP +"¡ƒ ›M»P phosphate-buffered saline (PBS: 137 mM NaCl, 2.7 mM KCl, 1.5 mM KH_2PO_4, pH 7.2)¡ A ¥ H Ø¿h:4æ/S/æ¤ , m /"S ª ¿ i " /‹ ª¿i 10 / ˜`¡A¯ ¥H13,000g "/" +t´ / 30 / ˜`¡A¤ œ / W h¥fl ƒ" …h(/ S "‹ fat cake)¡A ¤^»PS ¶+0.025 % Triton-X 100 /SPBS † ¥X¡A• nœ10 / ˜`¥H ¥h† ›M‰//Sfl ‰ Ł¤ ¥ ¡Af Æ,¥H13,000g "/" +t´ / 30 min¡A¤ œ / W h¥fl ƒ" …h¡AS Y ¥ i " ¤B† ›M»" "/fl " "w¡A¶ i / @ ¤ B S¥/„"qRa¤ " ¥+‰Ł¡C

¡]6¡^„q"a(SDS-PAGE)

/ W › ¥¿(¥'… , g ^ "«)¤°¿Eg ^ ˜ ¤«¿Æ/ ´ ' – o " " fl " "w/ ´ ' –o""fl " "w¡ Æ H0Œgl»PBio-Rad protein assay dye † V ƒ S@«¥Æ¥H/ ¥ œ ¥ œ « p ƒ595nm / Ų œ–o† J ¥ ‰Ł«¿@ C ¤ l ¥ [ƒ sample buffer (St 2% SDS, 15% glycerol, 1.5

M β-mercaptoethanol ／STris-HCl, pH 6.8)¡ Æb95°C ／ ／ ¥…5 ／ ˜`¡A¥H10,000 rpm ´ ＋ ť ｲ／ ˜`¤ ˇ fl " ‰／¡Œfl Bæ«Æ U … h S t ＋ ¥ ¡æŒH»"＄ ¡＃¥ ˇ ƒ "0.$t% SDS "" 10 % polyacrylaminde gel ¶ i ƒ „q¡"Æ– N £ƒP ／ ¶q" " ＋ ¥ ‰Ł´ ¡AƒA¥H Commassie Blue R-250 ‹ ¥ƒ ¡A‹ æß／ £P＋¡ƒ " " fl"＋ ›Mƒb^ ˜ " « ®¿Æ«Æefl " ‴w»– ＋ J ¥‰ŁS†SƒP¡C

3. †"G»P Q‰

– q ／ j ¥ „ « ˜ , A ／ W ／ Ł ¤ œ ¥ X " " ／ " ¯ ƒ fl " „ ¡A／ ´ ¥X¿W¥ ""fl " isoproterenol ¤ ¿E¡A o † {H^ "«¿ ® –q0¡B3¡Bl0¡B30 µM ＋ v " ¥W¥Æ¤ C † @ ／＋ ƒ (ml)" " fl "†¤O›Mƒi (h)˜ 'æ¥ X"· ´ fl"» ˜ S t ¶ｲ–]q•0.275¡B0.300¡B0.325¡Bl.075 mM ＋ v " ¥W¥¡[ƒ„1 ' ¥¡C^ "«isoproterenol ¥ D › n • » , ' ¥†¡ZÆƒ„ƒ]Q‰S¥／ ¤ˇ^ "«¿ ® ‹ 0¡ A † ›M · ƒ ＋ ‴¥» ´ fl"» ˜ ˜ 'æ¡¶C¥ ¥~¡ A ¥ Ł˜Ùokadaic acid ¤° ¿Efi ¡A／] o † { H ¤°¿Ł0¡®3¡Bl0¡B30 µM ＋ ¥／W／ fi ¡A¤ C † @ ／＋(mƒ)" fl " † ›MC／ ƒi (h)˜ 'æ¥ X"· ´ fl"» ˜ S ¶q–q0.100¡B0.900¡Bl.000¡Bl.075 mM ＋v " ¥ W¥¡[ƒ„2 ' ¥¡C¥ ' ^ "«okadaic acid ‹ O• » , 'DMSO ／¡Aƒ ƒ„• ^ "«¿ ® ‹ 0fi ¡A‹ O¥HDMSO ¥ N ¡A¥i ф{DMSO ＋B†zƒ ¥G• |S ¤ fl " † ›M"¥»· ´ fl " »˜"˜ 'æ¡O› L ＋ ¥ fŁU " „ ¸／ ´ ¥ X"ƒŁg^""＋¤ ¿Æ«Æ˜ 'æ" " fl"» ˜ S¶q , ß ¡]data not shown ¡ˆC／æ, ß„ˇ／®»P„ˇ／Gfi ／]¥i¥H ф{okadaic acid SC¿ ® ""¤ ¿E«K ¥i¥H„ Æ , ß／ j ‴fŁ‰Ł／ , ¡C

, g–˜¤ œ ／ ´fl"w" "Łƒ¡ ÆHSDS-PAGE „ ¤a／Łƒ¡／ " R " ›–¥ƒ‰Ł"˜ ／ˇ¡A¯ª ¥ ／ " ¯ ﬂ ‴w" →＋¥J ‰Łƒbˆ "«isoproterenol »Pokadaic acid fi ‰ T ŒS‡e{／ £P"˜"＋J¥ ‰Ł«‹ƒ¡] ƒ„3 ' ¥¡C¥ ¿ ¬ Ł "«¤ ¿E"˜„ •†¡ A – q fl " ／＋／›M ´ ¥ X"˜ﬂw¤" " →＋¥J ‰ŁS ¶q„ ß ƒh"˜ƒ＋72¡B56 »P32 kDa ¥＋J¥ ‰Ł¡]lane 1¡ˆF• ¥H30 µM "" isoproterenol ¤ ¿E«Æ ф{¤ ／／56 kDa "˜"＋J¥ ‰Ł« ‰œ¯ª• ／¡]lane 2¡ˆƒ • ¥H30 µM "" okadaic acid ¤ ¿E«Æ«h ф{¤ ／／56 kDa "˜"＋J¥ ‰Ł'œ¯ª... W [¡Aƒ 72 »P32 kDa ¥＋J¥ ‰Ł« ‰œ¯ª• ／ ¡]lane 3¡ˆC

¥ «／ƒw＋＋\ƒh／ m̃ˆ„Œ¡ A ˆ"¤« ¿E«e«Æ＋\ƒh" " fl"˜"w" →＋¥J ‰Ł"˜S ¶q›P" ^ • |†£¥˝S ¯ (Brasaemle et al., 2004)¡ŒPerilipin " ″ ／l¶q‹ ø« 56 kDa¡Aƒbepinephrine ¤ ¿E«Æƒ＋＋ ´ fl"˜"w" ›–"˜– $¸¡Aƒ„†{¶H»PS › "˜" ф{"˜"56 kD ＋J¥ ‰Ł' °‰Ł‹ "æ¡C ‹ OS ·N‹ Operelipin ¥H／ ¸¥ ¥~"˜"72 »P32 kDa "˜"＋J¥ ‰Ł«h» S¶ｲi@¤B„Œ ^ 'œ¡C

ƒ ／ ˜m¡G

Bialojan C. and Takai A. (1988) Inhibitory effect of a marine-sponge toxin, okadaic acid, on protein phosphatases. Specificity and kinetics. Biochemical Journal. 256:283-290

Bjorntorp P. (1985) Obesity and the risk of cardiovascular disease. Annals of Clinical Research. 17:3-9

Brasaemle D.L., Barer T., Wolins N.E., Serrero G., Blanchette M.J. and Londos C (1997)

Adipose differentiation-related protein is an ubiquitously expressed lipid storage droplet-associated protein. Journal of Lipid Research. 38: 2249-2263.
Brasaemle D.L., Dolis G., Shapiro L. and Wang R (2004) Proteomic analysis of proteins associated with lipid droplets of basal and lipolytically stimulated 3T3-L1 adipocytes. The Journal of Biological Chemistry. 279: 46835-46842.
Clifford G.M., McCormick DK, Londos C., Vernon R.G. and Yeaman S.J. (1998) Dephosphorylation of perilipin by protein phosphatases present in rat adipocytes. FEBS Letters. 435:125-129.
Fong T.H., Wang S.M. and Lin H.S. (1996) Immunocytochemical demonstration of a lipid droplet specific capsule in cultured Leydig cells of the golden hamsters. Journal of Cellular Biochemistry. 63: 366-373.
Fong T.H., Wu C.H., Liao E.W., Chang C.Y., Pai M.H., Chiou R.J. and Lee A.W. (2001) Association of globular beta-actin with intracellular lipid droplets in rat adrenocortical cells and adipocytes. Biochemical and Biophysical Research Communication. 289:1168-1174.
Greenberg A.S., Egan J.J., Wek S.A., Garty N.B., Blanchette M.J. and Londos C (1991) Perelipin, a major hormonally regulated adipocyte-specific phosphoprotein associated with the periphery of lipid storage droplets. The Journal of Biological Chemistry. 266: 11341-11346.
Hall P.F., Osawa S. and Mrotek J.J. (1981) The actions of various cytochalasins on mouse adrenal tumor cells in relation to trophic stimulation of steroidogenesis. Biochemica Biophysica Acta 676:338-344.
Haystead T.A., Sim A.T., Carling D., Honnor R.C., Tsukitani Y. Cohen P. and Hardie D.G. (1989) Effects of the tumor promoter okadaic acid on intracellular protein phosphorylation and metabolism. Nature. 337:78-81.
Leibel RL. Edens NK. Fried SK. (1989) Physiologic basis for the control of body fat distribution in humans. Annual Review of Nutrition. 9:417-43.
Londos C., Brasaemle D.L., Gruia-Gray J., Servetnick D.A., Schultz C.J., Levin D.M. and Kimmel A.R. (1995) Perilipin: unique proteins associated with intracellular neutral lipid droplets in adipocytes and steroidogenic cells. Biochemical Society Transactions. 23: 611-615.
Londos C., Brasaemle D.L., Schultz C.J., Segrest J.P. and Kimmel A.R. (1999) Perilipin, ADRP and other proteins that associate with intracellular neutral lipid droplets in animal cells. Seminars in Cell & Development Biology 10: 51-58.
Morimoto C., Tsujita T. and Okuda H. (1997) Norepinephrine-induced lipolysis in rat fat cells from viseral and subcutaneous sites: role of hormone-sensitive lipase and lipid droplets. Journal of Lipid Research 38: 132-138.
Morimoto C., Kiyama A., Kameda K., Ninomiya H., Tsujita T. and Okuda H. (2000) Mechanism of the stimulatory action of okadaic acid on lipolysis in rat fat cells. Journal of Lipid Research. 41:199-204.
Mrotek J.J. and Hall P.F. (1977) Response of adrenal tumor cells to adrenocorticotropin: site of inhibition by cytochalasin B. Biochemistry 16:3177-3181
Okuda H., Tsujita T. and Kinutani M. (1986) Studies on a protein kinase inhibitor-insensitive, phospholipase C-sensitive pathway of lipolysis in rat adipocytes. Pharmacological Research Communications. 18:877-93.
Olsson H. and Belfrage P. (1987) The regulatory and basal phosphorylation sites of

hormone-sensitive lipase are dephosphorylated by protein phosphatase-1, 2A and 2C but not by protein phosphatase-2B. European Journal of Biochemistry. 168: 399-405.
Rodbell M. (1964) Metabolism of isolated fat cells. The Journal of Biological Chemistry. 239: 375-380.
Servetnick D.A., Brasaemle D.L., Gruia G.J., Kimmel A.R., Wolff J. and Londos C. (1995) Perilipins are associated with cholesterol ester droplets in steroidogenic adrenal cortical and Leydig cells. The Journal of Biological Chemistry. 79: 16970-16973.
Wang S.M. and Fong T.H. (1995) A lipid droplet-specific capsule is present in rat adrenal cells: evidences from a monoclonal antibody. Biochemical & Biophysical Research Communication. 217: 81-88.
Yeaman S.J., Smith G.M., Jepson C.A., Wood S.L. and Emmison N. (1994) The multifunctional role of hormone-sensitive lipase in lipid metabolism. Advances in Enzyme Regulation. 34: 355-370.

„ ˇ/ø»Þ» ¡'œ

1

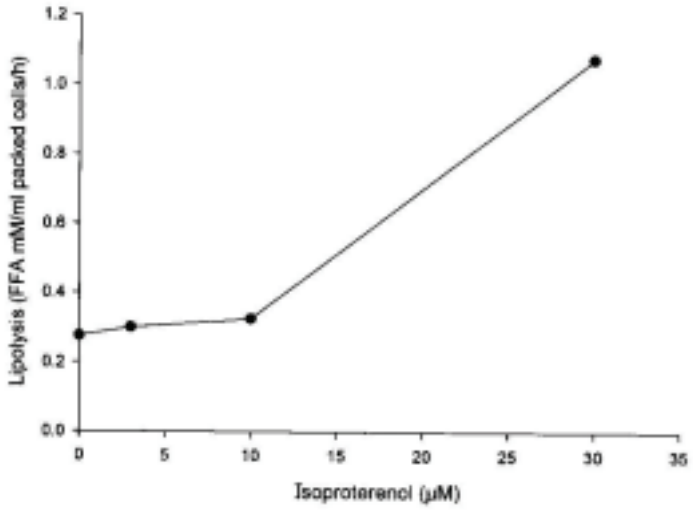

Fig.1 Effect of isoproterenol on lipolysis in rat fat cells.

2

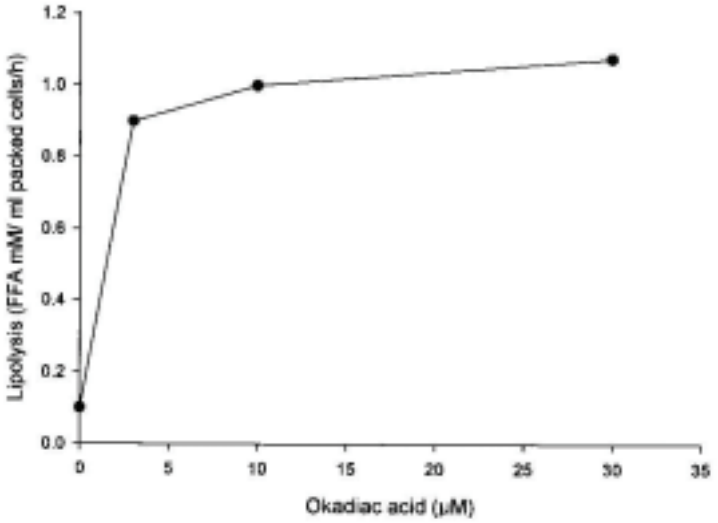

Fig. 2 Effect of okadaic acid on lipolysis in rat fat cells.

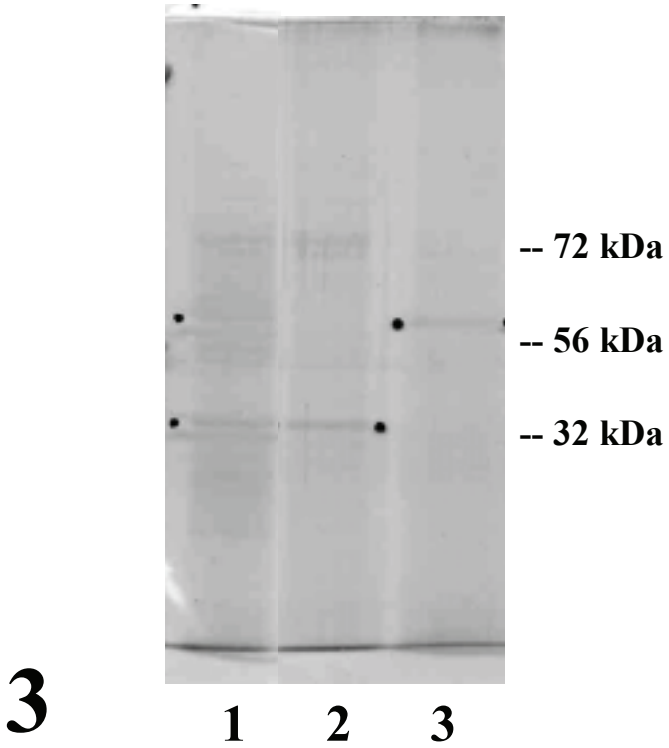

Fig. 3 SDS-PAGE of proteins of lipid droplet fraction from rat fat cells. Lane 1: normal fat cells. Lane 2: isoproterenol-stimulated fat cells. Lane 3: okadaic acid-stimulated fat cells. Note that the 56 kDa protein decreased during isoproterenol treatment but it increased during okadaic acid treatment.