RAPID SUBMISSION

ALTERATIONS OF SERUM BRAIN-DERIVED NEUROTROPHIC FACTOR LEVELS IN EARLY ALCOHOL WITHDRAWAL

MING-CHYI HUANG^{1,2,5}, CHUN-HSIN CHEN^{2,3}, CHIA-HUI CHEN⁴, SHING-CHENG LIU^{1,2}, CHIA-JEN HO¹, WINSTON W SHEN^{2,3*} and SY-JYE LEU^{5*}

¹Department of Psychiatry, Taipei City Psychiatric Center, Taipei City Hospital, Taipei, Taiwan ²Department of Psychiatry, School of Medicine, Taipei Medical University, Taipei, Taiwan ³Department of Psychiatry, Taipei Medical University-Wan Fang Hospital, Taipei, Taiwan ⁴Division of Mental Health and Substance Abuse Research, National Health Research Institute, Taiwan ⁵Institute of Cell and Molecular Biology, Taipei Medical University, Taipei, Taiwan

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Abstract — **Aims:** Alcohol withdrawal-enhanced neurotoxicity contributes to the addictive process. Brain-derived neurotrophic factor (BDNF) plays an important role in neuronal plasticity and learning. In this study, we explored the changes of serum BDNF levels in alcoholic patients at baseline and after one-week alcohol withdrawal. **Methods:** Twenty-five alcoholic patients were admitted for alcohol detoxification treatment, and 22 healthy control subjects were also enrolled. We collected blood samples of the patient group on the first and seventh day of alcohol withdrawal, and measured serum BDNF level with sandwich enzyme-linked immunosorbent assay. The severity of withdrawal symptoms was evaluated by the Clinical Institute Withdrawal Assessment-Alcohol, Revised every eight hours. **Results:** Serum BDNF levels did not differ significantly between alcoholic patients and control subjects. But BDNF levels were found to be significantly increased one week after alcohol withdrawal (from 13.9 ± 3.8 ng/ml to 15.4 ± 3.8 ng/ml, P = 0.03). **Conclusions:** The present study suggests that elevated serum BDNF levels were found in early alcohol withdrawal, implying that BDNF may involve in neuroadaptation during the period.

INTRODUCTION

Brain-derived neurotrophic factor (BDNF) is the most abundant member of the nerve growth factor (NGF)-related family of neurotrophic factors in brain, and responsible for neuronal survival, outgrowth, and differentiation during development (Snider, 1994; Lewin and Barde, 1996). In adults, BDNF has also been found to serve as a neurotransmitter modulator to participate in neuronal plasticity, such as long-term potentiation and learning. (Hyman et al., 1991; Thoenen, 1995; Li et al., 1998; Lyons et al., 1999; Hall et al., 2000; Guillin et al., 2001). Although BDNF is highly concentrated in the nervous system, it is also present in human and rat peripheral blood, and is more concentrated in the serum. Previous studies suggest that BDNF can cross the blood-brain barrier (Pan et al., 1998), that serum BDNF levels may reflect BDNF levels in the brain (Radka et al., 1996; Karege et al., 2002), and that these levels are relatively stable among adult primates including humans (Mori et al., 2003).

Activity-dependent activation of BDNF is associated with the neuroadaptation, which is involved in the development of addiction (Horger *et al.*, 1999; Guillin *et al.*, 2001; Hall *et al.*, 2003; McGough *et al.*, 2004). BDNF is responsible for normal expression of dopamine D3 receptor in nucleus accumbens where it receives mesolimbic projections from ventral tegmental area, involving in reward system (Guillin *et al.*, 2001). Increased BDNF expression has been reported to be associated with drug abuse, such as morphine (Numan *et al.*, 1998), amphetamine (Meredith *et al.*, 2002), cocaine (Grimm *et al.*, 2003), and delta-tetrahydrocannabinol (Butovsky *et al.*, 2005).

Alcohol dependence is a chronic disease characterized by uncontrolled drinking and a chronically relapsing course. Several lines of evidence suggest that BDNF also plays some role in developing alcohol addiction. The finding from the in vitro study showed biphasic changes, that BDNF mRNA expression is increased under acute alcohol exposure, but decreased after continuous ethanol exposure (McGough et al., 2004). In animal studies, BDNF mRNA expression has been found to be reduced following chronic alcohol exposure, but increased after its withdrawal (MacLennan et al., 1995; Tapia-Arancibia et al., 2001). Compared to wild-type mice, heterozygous BDNF (+/-) mutant mice can self-administer a larger amount of alcohol (Hensler et al., 2003). The adaptive changes in the brain during alcohol withdrawal which contribute to the progressive nature of alcohol dependence (Koob, 2003) are associated with neurodegeneration (Littleton et al., 2001; Crews et al., 2004). Therefore, neurotrophic factors for those adaptive changes might play an important role during alcohol withdrawal.

To date, only one study exploring neurotrophin during alcohol withdrawal in human showed a significant increase in plasma NGF level (Aloe *et al.*, 1996). We assumed both that alcohol withdrawal and its neurotoxic effect could induce a neuroadaptative response and that the withdrawal symptoms might reflect central excitotoxicity (Tsai and Coyle, 1998; Littleton *et al.*, 2001). Therefore, we carried out this study to test the hypothesis that serum BDNF in alcoholic patients would be increased after alcohol withdrawal and to explore the correlation between BDNF and the severity of alcohol withdrawal symptoms.

^{*}Author to whom correspondence should be addresses: Institute of Cell and Molecular Biology, Taipei Medical University, Taipei, Taiwan. Tel: +886 2 27361661 ext 3414; Fax: +886 2 23778620; E-mail: cmbsycl@tmu.edu.tw.*Winston W. Shen as the co-corresponding author for equivalent contribution to the article.

METHODS

Alcoholic subjects were recruited from an alcohol detoxification treatment unit in Taipei City Psychiatric Center. Patients were invited to participate in the study if they fulfilled the following inclusion criteria of age being between 20 to 60 years, being admitted for alcohol detoxification, and having DSM-IV diagnosis of alcohol dependence. Excluded were those who were co-morbid with other current non-nicotine substance abuse or dependence; had significant physical illnesses, such as ischemic heart disease or poorly controlled diabetes mellitus; had other psychiatric disorders, such as schizophrenia bipolar disorder, or major depressive disorder; had been treated with antipsychotics or antidepressant; and suffered from severe cognitive impairment with difficulty in understanding the study content. We assessed the patients with initial clinical interview to ascertain the DSM-IV diagnoses. Then the subjects received physical examination and urine toxicology test to screen for illicit drugs and to exclude other substance use disorders. Approval was obtained from the institutional review board at Taipei City Psychiatric Center before the study began. After initial assessments, we gave all eligible patients a comprehensive description of the study and then recruited them into the study project after obtaining written informed consent for participation.

We collected subjects' socio-demographic data (age, gender, educational level, and marital status). Then we interviewed patients to gather drinking history (age at first intoxication, age at dependence, and average daily amount of alcohol consumption in the past one month). Alcohol consumption was stopped abruptly and completely at admission and patients' withdrawal symptoms were evaluated using the Clinical Institute Withdrawal Assessment-Alcohol, Revised (CIWA-Ar) (Sullivan et al., 1989) every eight hours by trained nurses. Average of CIWA-Ar score and the highest CIWA-Ar score in the first day were recorded as the baseline withdrawal severity. The patients received fixed-dose schedule of alcohol detoxification treatment with oral lorazepam 2 mg for four doses with gradual tapering thereafter, and the as-needed use of oral trazodone (50 mg) at night for sleeping problems. They also received multivitamins. All smoking patients simultaneously received aided nicotine patches, with dose administration adjusted by individual's average smoking amount, for smoking cessation program.

The control group included healthy subjects without known physical and psychiatric illnesses identified in the interview and the results of routine laboratory tests for liver and renal functions. They did not meet the diagnostic criteria of alcohol abuse or dependence in the past nor drink alcohol during the previous three months.

Serum samples of the study inpatients were collected on the next morning of admission for detoxification treatment (baseline) and on the seventh day following the treatment (one week). The serum samples were stored at -80° C until used for the assay. Serum BDNF levels were measured using the BDNF Emax Immunoassay System Kit according to the manufacturer's instructions (Promega, USA). We measured serum BDNF levels of all subjects on the same day.

The results are presented as mean and standard deviation (SD). With independent *t*-test, we analyzed the differences between groups. With paired *t*-test, we analyzed the differences

of serum BDNF level collected at baseline and one week after alcohol withdrawal. Pearson's correlation test was used to determine the correlation between BDNF and clinical parameters. The differences between the groups were considered significant, if *P*-values were smaller than 0.05.

RESULTS

A total of 25 patients (21 male and 4 female) and 22 healthy controls (19 male and 3 female) were enrolled, with mean age \pm SD being 41.3 \pm 7.8 and 43.6 \pm 6.3 years, respectively. In the alcoholic patients, the average duration of alcohol dependence was 8.5 \pm 5.8 years, and the average daily amount of alcohol consumption in the past one month was 208.8 \pm 125.4 grams of pure ethanol. The classical serum biomarkers (mean \pm SD) of drinking in our patients were: AST 122.0 \pm 101.2 U/I, ALT 54.1 \pm 26.6 U/I, gamma-glutamyltransferase (GGT) 693.57 \pm 762.1 U/I, MCV 95.0 \pm 9.7 fl, and uric acid 7.4 \pm 2.0 mg/dl (n = 22).

Figure 1 showed the serum BDNF levels of alcoholic patients in two different time points and normal controls. Serum BDNF levels of the alcoholic group were significantly increased from 13.9 ± 3.8 ng/ml at baseline to 15.4 ± 3.8 ng/ml after oneweek alcohol withdrawal (P = 0.03). Despite neither of these values differed significantly from that of the healthy control group, 15.8 ± 3.7 ng/ml, there was a nonsignificant trend that the baseline BDNF levels of alcoholic patients were lower than those of control subjects (P = 0.09). The magnitude of oneweek BDNF elevation did not significantly correlate with clinical characteristics (age, amount of cigarette consumption, age at first alcohol intoxication, age at alcohol dependence, and duration of alcohol dependence), but had marginally significant positive correlation with average amount of alcohol consumption (r = 0.38, P = 0.06).

A significant correlation was also found between baseline BDNF levels and the highest first-day CIWA-Ar scores (r = 0.44, P = 0.03) or average first-day CIWA-Ar scores (r = 0.45, P = 0.03) (Fig. 2). Even after controlling the amount of cigarettes, the correlation between baseline BDNF levels and average of first-day CIWA-Ar scores was still significant (r = 0.46, P = 0.02). The other clinical variables (age, age at first alcohol intoxication, age at alcohol dependence, duration of alcohol dependence, average daily amount of alcohol consumption, or biochemical values) were not correlated with their baseline BDNF levels.

DISCUSSION

This is the first study to show significant increase in serum BDNF levels after one-week alcohol withdrawal in patients with alcohol dependence, and a significant positive correlation between patients' baseline BDNF levels and their baseline withdrawal severity. In addition, the results of this study revealed a trend that baseline BDNF levels of alcoholic patients were lower than those of control subjects, but BDNF levels after one-week alcohol withdrawal were approximately equivalent to those in controls. Despite nicotine can also increase BDNF expression (Le Foll *et al.*, 2005), baseline BDNF level still remained significantly correlated with baseline withdrawal severity even after controlling cigarette amount

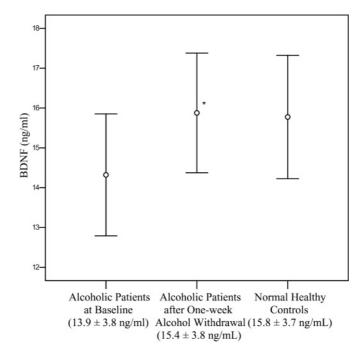


Fig. 1. Serum BDNF levels (mean \pm SD) of alcohol patients (n = 25) at baseline and after one-week alcohol withdrawal as well as of the healthy controls (n = 22). *Significant difference (P = 0.03) between BDNF levels after one-week alcohol withdrawal and those at baseline of alcoholic patients.

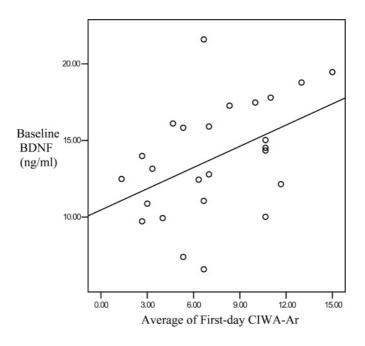


Fig. 2. The correlation between baseline serum BDNF levels and the average of first-day CIWA-Ar scores in alcoholic patients (n = 25) (r = 0.45, P = 0.03).

statistically. Interestingly, a near-significant trend existed between the magnitude of BDNF elevation and the amount of alcohol consumption.

The finding of increased neurotrophin after alcohol withdrawal in our study is consistent with a previous human study showing increased NGF after alcohol withdrawal (Aloe *et al.*, 1996). In animal study, the pattern of alcohol administration might influence BDNF expression. Repeated episodic alcohol exposure, to exhibit withdrawal phenomenon, caused BDNF and NGF elevation in hippocampus, while chronic alcohol exposure leads to decreased BDNF expression in rats(Miller, 2004). Following alcohol withdrawal after four-weeks alcohol vapor inhalation, a significant increase in BDNF mRNA expression was found in rat hippocampus and hypothalamus (Tapia-Arancibia *et al.*, 2001). The disinhibition of BDNF could exert protective function against neuronal damage and stimulation of sprouting and synaptic reorganization during withdrawal from chronic alcohol ingestion (Tapia-Arancibia

et al., 2001). Phosphorylated cAMP response element bindingprotein (CREB) regulates the expression of several downstream cAMP-inducible genes, including BDNF. Phosphorylated CREB protein levels in hippocampus increased after alcohol withdrawal after binge alcohol treatment (Bison and Crews, 2003), possibly contributing to increased neurogenesis during abstinence (Nixon and Crews, 2004).

Sustained stress, kindling-induced seizures, head trauma, or hypoglycemic coma can trigger elevated BDNF expression in the CNS of rats (Lindvall et al., 1994). Glutamate release and calcium influx were postulated to be the key factors responsible for increasing BDNF during the brain insults. Thus, neuronal excitotoxicity mediated by glutamate receptors has long been implicated in the pathophysiology of alcohol withdrawal phenomenon and resulting neuronal damage (Tsai and Coyle, 1998; Littleton et al., 2001). BDNF treatment in neuron culture was shown to prevent NMDA excitotoxicity sensitized by ethanol (Crews et al., 2004). Therefore, BDNF could be involved in promoting the resilience of brain cells and needs to be elevated to cope with aggressive stress, such as NMDA overexcitation during alcohol withdrawal. The result of an animal study showed that compensatory neurogenesis, reflected by increased cell proliferation, was correlated with alcohol withdrawal severity despite the use of diazepam (10 mg/day) (Nixon and Crews, 2004). Consistent with these previous findings, our study showed a significant positive correlation between baseline BDNF level and baseline withdrawal severity, supporting a neuomodulating role of BDNF to oppose withdrawal-enhanced neurotoxicity.

Regarding BDNF gene polymorphism, alcoholic subjects with a history of delirium tremens had a higher frequency of AA genotypes and A allele, which has been peculated to correlate with impaired BDNF secretion (Matsushita et al., 2004). Thus, those with inadequate BDNF expression to counteract the neurotoxicity could experience more severe withdrawal. BDNF levels in mesolimbic dopamine system progressively elevated after cocaine withdrawal. The prolonged BDNF increase was implicated in neuroadaptation of craving (Grimm et al., 2003). Moreover, the volume of alcohol intake was also reported to be associated with craving during alcohol withdrawal (Hillemacher et al., 2006). These findings might, in part, explain our intriguing clinical finding of correlation with near-significant correlation of magnitude of one-week post-withdrawal BDNF increment with the amount of alcohol consumption.

Plasminogen activator inhibitor 1 has been suggested to inhibit the action of tissue-type plasminogen activator (tPA), which cleaves plasminogen and thus, activates plasmin. Plasmin proteolytically cleaves pro-BDNF to yield mature BDNF (Pang et al., 2004). The result of a previous study indicated that PAI-1 has been decreased significantly from day 1 to day 22 after alcohol withdrawal (Delahousse et al., 2001). Similarly, the finding of another study showed that PAI-1 levels were significantly higher in alcoholics at baseline and decreased after one-week alcohol withdrawal (Soardo et al., 2006). Moreover, an animal study showed that tPA activity is upregulated to promote stress-induced neuronal remodeling in mouse brain under acute restraint stress (Pawlak et al., 2003). Thus, we postulate that attenuated PAI-1 after alcohol withdrawal may result in increased tPA activity, which further enhances higher BDNF levels as an adaptation to the physiological stress. The finding of elevated BDNF in our patients after alcohol withdrawal is in agreement with this supposed mechanism.

Generalization of the research data should be cautious because this study had three limitations. First, the sample size was small. The difference between baseline BDNF levels in alcoholic patients and controls was nearly significant (P = 0.09). Therefore, larger samples are needed to examine the BDNF downregulation in alcoholic patients. Second, we could not rule out the effects of medications, such as lorazepam on BDNF levels in our alcoholic patients. But the pertinent report still remains paucity. The only indirect clue is that acute or chronic injection of another benzodiazepine, chlordiazepoxide did not protect neuronal damage in animals (Haynes et al., 2004). Thus, the influence of lorazepam on BDNF levels in our patients might have been minimal, but still needs further clarification. Third, the effect of cigarette smoking cessation on BDNF level might confound the results in this study. A recent report indicated that plasma BDNF levels in chronic smokers increased following two-month unaided smoking cessation (Kim et al., 2007). Although we gave aided nicotine patch to minimize the confounding effect of smoking cessation on BDNF elevation during one-week interval, however, we still cannot rule out the potential impact. Adequate subjects to examine the difference of BDNF levels in both smokers and nonsmokers of alcoholic patients during withdrawal are needed.

In conclusion, this study showed that BDNF levels did not differ between alcoholic patients and controls, but increased after one-week alcohol withdrawal in alcoholic patients. Furthermore, the baseline BDNF level was found positively correlated with baseline alcohol withdrawal severity. These findings support that BDNF is involved in the neuroadaptation of alcohol withdrawal, a stress condition or brain insult which requires neurotrophin to promote neuronal resilience or survival.

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REFERENCES

- Aloe, L., Tuveri, M. A., Guerra, G. et al. (1996) Changes in human plasma nerve growth factor level after chronic alcohol consumption and withdrawal. Alcoholism-Clinical and Experimental Research 20, 462–465.
- Bison, S. and Crews, F. (2003) Alcohol withdrawal increases neuropeptide Y immunoreactivity in rat brain. *Alcoholism-Clinical and Experimental Research* 27.
- Butovsky, E., Juknat, A., Goncharov, I. et al. (2005) In vivo upregulation of brain-derived neurotrophic factor in specific brain areas by chronic exposure to delta-tetrahydrocannabinol. Journal of Neurochemistry 93, 802–811.
- Crews, F. T., Collins, M. A., Dlugos, C. et al. (2004) Alcohol-induced neurodegeneration: When, where and why? Alcoholism-Clinical and Experimental Research 28, 350–364.
- Delahousse, B., Maillot, F., Gabriel, I., Schellenberg, F., Lamisse, F. and Gruel, Y. (2001) Increased plasma fibrinolysis and tissue-type

plasminogen activator/tissue-type plasminogen activator inhibitor ratios after ethanol withdrawal in chronic alcoholics. Blood Coagulation and Fibrinolysis **12**, 59–66. Grimm, J. W., Lu, L., Hayashi, T. Hope, B. T., Su, T. P. and Shaham,

- Y. (2003) Time-dependent increases in brain-derived neurotrophic factor protein levels within the mesolimbic dopamine system after withdrawal from cocaine: Implications for incubation of cocaine craving. The Journal of Neuroscience 23, 742-747.
- Guillin, O., Diaz, J., Carroll, P., Griffon, N., Schwartz, J. C. and Sokoloff, P. (2001) BDNF controls dopamine D3 receptor expression and triggers behavioural sensitization. Nature 411, 86-89.
- Hall, F. S., Drgonova, J., Goeb, M. and Uhl, G. R. (2003) Reduced behavioral effects of cocaine in heterozygous brain-derived neurotrophic factor (BDNF) knockout mice. Neuropsychopharmacology 28, 1485–1490.
- Hall, J., Thomas, K. L. and Everitt, B. J. (2000) Rapid and selective induction of BDNF expression in the hippocampus during contextual learning. Nature Neuroscience 3, 533–535.
- Haynes, L. E., Barber, D. and Mitchell, I. J. (2004) Chronic antidepressant medication attenuates dexamethasone-induced neuronal death and sublethal neuronal damage in the hippocampus and striatum. Brain Research 1026, 157–167.
- Hensler, J. G., Ladenheim, E. E. and Lyons, W. E. (2003) Ethanol consumption and serotonin-1A (5-HT1A) receptor function in heterozygous BDNF (±) mice. Journal of Neurochemistry 85, 1139-1147.
- Hillemacher, T., Bayerlein, K., Wilhelm, J. et al. (2006) Volume intake and craving in alcohol withdrawal. Alcohol and Alcoholism 41, 61-
- Horger, B. A., Iyasere, C. A., Berhow, M. T., Messer, C. J., Nestler, E. J. and Taylor, J. R. (1999) Enhancement of locomotor activity and conditioned reward to cocaine by brain-derived neurotrophic factor. The Journal of Neuroscience 19, 4110-4122
- Hyman, C., Hofer, M., Barde, Y. A. et al. (1991) BDNF is a neurotrophic factor for dopaminergic neurons of the substantia nigra. Nature 350, 230-232.
- Karege, F., Schwald, M. and Cisse, M. (2002) Postnatal developmental profile of brain-derived neurotrophic factor in rat brain and platelets. Neuroscience Letters 328, 261-264.
- Kim, T. S., Kim, D. J., Lee, H. and Kim, Y. K. (2007) Increased plasma brain-derived neurotrophic factor levels in chronic smokers following unaided smoking cessation. *Neuroscience Letters* **423**, 53-57.
- Koob, G. F. (2003) Alcoholism: Allostasis and beyond. Alcoholism-Clinical and Experimental Research 27, 232-243.
- Le Foll, B., Diaz, J. and Sokoloff, P. (2005) A single cocaine exposure increases BDNF and D3 receptor expression: Implications for drugconditioning. *Neuroreport* **16**, 175–178. Lewin, G. R. and Barde, Y. A. (1996) Physiology of the neurotrophins.
- Annual Review of Neuroscience 19, 289–317
- Li, Y. X., Zhang, Y., Lester, H. A., Schuman, E. M. and Davidson, N. (1998) Enhancement of neurotransmitter release induced by brainderived neurotrophic factor in cultured hippocampal neurons. The Journal of Neuroscience 18, 10231-10240.
- Lindvall, O., Kokaia, Z., Bengzon, J., Elmer, E. and Kokaia, M. (1994) Neurotrophins and brain insults. Trends in Neurosciences 17, 490-496
- Littleton, J. M., Lovinger, D., Liljequist, S., Ticku, R., Matsumoto, I. and Barron, S. (2001) Role of polyamines and NMDA receptors in ethanol dependence and withdrawal. Alcoholism: Clinical and Experimental Research 25(5 suppl), 132S–136S.
- Lyons, W. E., Mamounas, L. A., Ricaurte, G. A. et al. (1999) Brainderived neurotrophic factor-deficient mice develop aggressiveness and hyperphagia in conjunction with brain serotonergic abnormali-

ties. Proceedings of the National Academy of Sciences of the United States of America 96, 15239–15244.

- MacLennan, A. J., Lee, N. and Walker, D. W. (1995) Chronic ethanol administration decreases brain-derived neurotrophic factor gene expression in the rat hippocampus. Neuroscience Letters 197, 105-108
- Matsushita, S., Kimura, M., Miyakawa, T. et al. (2004) Association study of brain-derived neurotrophic factor gene polymorphism and alcoholism. Alcoholism: Clinical and Experimental Research 28, 1609-1612.
- McGough, N. N., He, D. Y., Logrip, M. L. et al. (2004) RACK1 and brain-derived neurotrophic factor: A homeostatic pathway that regulates alcohol addiction. The Journal of Neuroscience 24, 10542-10552.
- Meredith, G. E., Callen, S. and Scheuer, D. A. (2002) Brain-derived neurotrophic factor expression is increased in the rat amygdala, piriform cortex and hypothalamus following repeated amphetamine administration. Brain Research 949, 218-227.
- Miller, M. W. (2004) Repeated episodic exposure to ethanol affects neurotrophin content in the forebrain of the mature rat. Experimental Neurology 189, 173-181.
- Mori, T., Shimizu, K. and Hayashi, M. (2003) Levels of serum brainderived neurotrophic factor in primates. Primates 44, 167-169.
- Nixon, K. and Crews, F. T. (2004) Temporally specific burst in cell proliferation increases hippocampal neurogenesis in protracted abstinence from alcohol. The Journal of Neuroscience 24, 9714–9722.
- Numan, S., Lane-Ladd, S. B., Zhang, L. et al. (1998) Differential regulation of neurotrophin and trk receptor mRNAs in catecholaminergic nuclei during chronic opiate treatment and withdrawal. The Journal of Neuroscience 18, 10700-10708.
- Pan, W., Banks, W. A., Fasold, M. B., Bluth, J. and Kastin, A. J. (1998) Transport of brain-derived neurotrophic factor across the blood-brain barrier. Neuropharmacology 37, 1553-1561.
- Pang, P. T., Teng, H. K., Zaitsev, E. et al. (2004) Cleavage of proBDNF by tPA/plasmin is essential for long-term hippocampal plasticity. Science **306**, 487–491.
- Pawlak, R., Magarinos, A. M., Melchor, J., McEwen, B. and Strickland, S. (2003) Tissue plasminogen activator in the amygdala is critical for stress-induced anxiety-like behavior. Nature Neuroscience 6, 168-174.
- Radka, S. F., Holst, P. A., Fritsche, M., and Altar, C. A. (1996) Presence of brain-derived neurotrophic factor in brain and human and rat but not mouse serum detected by a sensitive and specific immunoassay. Brain Research 709, 122-301.
- Snider, W. D. (1994) Functions of the neurotrophins during nervous system development: What the knockouts are teaching us. Cell 77, 627-638.
- Soardo, G., Donnini, D., Varutti, R. et al. (2006) Effects of alcohol withdrawal on blood pressure in hypertensive heavy drinkers. Journal of Hypertension 24, 1493–1498.
- Sullivan, J. T., Sykora, K., Schneiderman, J., Naranjo, C. A. and Sellers, E. M. (1989) Assessment of alcohol withdrawal: The revised clinical institute withdrawal assessment for alcohol scale (CIWA-Ar). British Journal of Addiction 84, 1353–1357.
- Tapia-Arancibia, L., Rage, F., Givalois, L., Dingeon, P., Arancibia, S. and Beauge, F. (2001) Effects of alcohol on brain-derived neurotrophic factor mRNA expression in discrete regions of the rat hippocampus and hypothalamus. Journal of Neuroscience Research **63**, 200–208.
- Thoenen, H. (1995) Neurotrophins and neuronal plasticity. Science 270, 593-598.
- Tsai, G. and Coyle, J. T. (1998) The role of glutamatergic neurotransmission in the pathophysiology of alcoholism. Annual Review of Medicine 49, 173–184.